



OPEN ACCESS

EDITED BY

Haiqiang Yao,
Beijing University of Chinese Medicine, China

REVIEWED BY

Antonietta G. Gravina,
University of Campania Luigi Vanvitelli, Italy
Bin Bao,
Boston Children's Hospital and Harvard
Medical School, United States

*CORRESPONDENCE

Ying Zhu

✉ zhuying089@126.com

Yin Xu

✉ 311118@hnu.edu.cn

[†]These authors have contributed equally to this work

RECEIVED 02 December 2024

ACCEPTED 10 February 2025

PUBLISHED 26 February 2025

CITATION

Long D, Mao C, Zhang W, Zhu Y and Xu Y (2025) Natural products for the treatment of ulcerative colitis: focus on the JAK/STAT pathway.
Front. Immunol. 16:1538302.
doi: 10.3389/fimmu.2025.1538302

COPYRIGHT

© 2025 Long, Mao, Zhang, Zhu and Xu. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Natural products for the treatment of ulcerative colitis: focus on the JAK/STAT pathway

Dan Long^{1†}, Chenhan Mao^{2†}, Wei Zhang¹, Ying Zhu^{1*} and Yin Xu^{1*}

¹Department of Gastroenterology, The First Hospital of Hunan University of Chinese Medicine, Changsha, Hunan, China, ²Affiliated Hospital of Integrated Traditional Chinese and Western Medicine, Nanjing University of Chinese Medicine, Nanjing, Jiangsu, China

Ulcerative colitis (UC) is an autoimmune disease with an incompletely understood pathogenesis. The Janus kinase (JAK)/signal transducer and activator of transcription (STAT) signaling pathway plays a key role in immune response and inflammation. More and more studies demonstrated that JAK/STAT signaling pathway is associated with the pathogenesis of UC. The JAK/STAT pathway affects UC in multiple ways by regulating intestinal inflammatory response, affecting intestinal mucosal barrier, modulating T cell homeostasis, and regulating macrophages. Encouragingly, natural products are promising candidates for the treatment of UC. Natural products have the advantage of being multi-targeted and rich in therapeutic modalities. This review summarized the research progress of JAK/STAT pathway-mediated UC. Furthermore, the latest studies on natural products targeting the JAK/STAT pathway for the treatment of UC were systematically summarized, including active ingredients such as arbutin, aloe polysaccharide, berberine, matrine, curcumin, Ginsenoside Rh2, and so on. The aim of this paper is to provide new ideas for drug development to regulate JAK/STAT signaling for treating UC.

KEYWORDS

Janus kinase (JAK), signal transducer and activator of transcription (STAT), ulcerative colitis, natural products, traditional herbal medicine

1 Introduction

Ulcerative colitis (UC) is a refractory digestive disease defined by recurring and remitting mucosal inflammation. Common clinical signs of UC include recurring stomach pain, diarrhea, and hematochezia. Typical medications used clinically as the primary treatment option for UC include aminosalicylates, corticosteroids, immunosuppressants, biological agents, and microecologics (1). Despite the large number of drugs available for the treatment of UC, its treatment remains complex and challenging due to a variety of side effects, medication tolerance, and high relapse rates (2). Therefore, further development of more effective treatments for UC has become urgent.

The Janus tyrosine protein kinase (JAK)/signal transducer and activator of transcription (STAT) signaling pathway has been identified as a classical inflammatory pathway. The JAK/STAT signaling is involved in biological processes such as cell proliferation, differentiation, and apoptosis. The JAK/STAT pathway plays a key role in the immune response and has become a focus of research in autoimmune and inflammatory diseases (3). Notably, the JAK/STAT pathway is associated with damage induced by exaggerated innate immune system response stimulated by immune checkpoint inhibitors (4). JAK/STAT signaling is frequently dysregulated in UC patients, indicating the importance of JAK/STAT regulation in UC (5, 6). Furthermore, in the colitis rat model, the severity of intestinal illness was positively associated to the expression of JAK2 and STAT3 (7). Theoretically, intervening in the JAK/STAT signaling pathway using safe and effective drugs may be an effective way to alleviate or treat UC. Currently, several JAK inhibitors have achieved efficacy in numerous clinical settings. The non-selective JAK inhibitor tofacitinib has been approved for moderate and severe UC (8). Encouragingly, natural products shows potential for the treatment of UC (9, 10). However, the existing studies are scattered and unsystematic. To our knowledge, this is the first thorough review that elaborates on recent advances of active ingredients in treating UC by modulating the JAK/STAT signaling pathway.

In this review, the current knowledge of the composition, activation, and regulation of the JAK/STAT pathway was discussed. Secondly, the role and mechanism of the JAK/STAT pathway in UC were particularly emphasized. Finally, we also systematically summarized the application of natural products targeting JAK/STAT signaling against UC. This review aims to provide new research ideas for traditional Chinese medicine (TCM) in the prevention and treatment of UC.

2 JAK/STAT pathway

2.1 Composition and activation of the JAK/STAT pathway

JAK is a non-receptor tyrosine protein kinase that is activated by numerous cytokines and initiates downstream target genes via STAT, which in turn regulates a variety of cellular functions (3). The JAK/STAT pathway consists of three main components, including tyrosine kinase-associated receptors, JAKs, and STATs. Four types of JAKs have been identified, including JAK1, JAK2, JAK3, and tyrosine kinase 2 (TYK2). Among them, JAK3 is expressed only in bone marrow and lymphocytes, while other members are widely found in various tissues and organs in the body (11). The JAK proteins are made up of FERM (the complex of four point one, ezrin, radixin, and moesin), Src homology domain (SH2), pseudokinase, and kinase domains. STAT proteins are downstream signaling molecules of JAK. STATs consist of seven members, namely STAT1, STAT2, STAT3, STAT4, STAT5A, STAT5B, and STAT6, which are widely distributed in various tissues. STATs proteins mainly contain five structural domains, including N-terminal conserved sequences, DNA-binding region,

Src homology domain 3 (SH3) structural region, SH2 structural region, and C-terminal transcriptional activation sequence. The SH2 structural area of STATs is identical to the analogous core sequence in JAKs, which is in charge of recognizing individual JAKs. Cytokines attach to cell-surface receptors, which dimerize and stimulate the polymerization and phosphorylation of JAKs. Activated JAKs can then bind to the SH2 structural domain of STATs, which are activated by phosphorylation modification and ultimately enter the nucleus as homodimers or heterodimers, thus promoting transcription of specific target genes (12). STAT is then dephosphorylated in the nucleus and returned to the cytoplasm (12). Among the STAT family, STAT3 has been recognized to play a central role in signaling from the plasma membrane to the nucleus (13). STAT3 is activated by phosphorylation of tyrosine (Y705) or serine (S727) residues in the transactivation domains, creating a STAT3 dimer that moves into the nucleus, where it promotes the transcription of target genes. Phosphorylation of STAT3 at the Y705 site occurs predominantly through members of the JAK family, whereas phosphorylation at the S727 site is usually carried out by mitogen-activated protein kinase, cell cycle protein-dependent kinase 5, and protein kinase C.

2.2 Negative regulation of the JAK/STAT pathway

The JAK/STAT pathway is primarily regulated negatively by three types of factors: suppressor of cytokine signaling (SOCS), protein inhibitor of activated STAT (PIAS), and protein tyrosine phosphatase (PTP) (14). (Figure 1). The SOCS family is the main signaling molecule that weakens the JAK/STAT pathway, including CIS, SOCS1, SOCS2, SOCS3, SOCS4, SOCS5, SOCS6, and SOCS7. Activated STAT entering the nucleus promotes the transcription of SOCS, which has a negative regulatory effect on JAK/STAT signaling by inhibiting STAT receptor binding, inactivating JAK through N-terminal kinase inhibition, or binding and ubiquitinating JAK or STAT for proteasomal destruction (15). PIAS can interact with STAT to prevent STAT dimerization or prevent STAT dimers from binding to DNA. PTP can dephosphorylate JAK by interacting with receptors as a phosphatase. It can also directly dephosphorylate STAT dimers to block JAK/STAT signaling transmission (16).

2.3 JAK/STAT pathway and melanocortin system

The melanocortin system is a complex signaling system composed of multiple hormones, neuropeptides, and receptors, which exerts a widespread regulatory role in the physiological and pathological processes in the body. α -, β - and γ -melanocyte-stimulating hormone (MSH) and adrenocorticotrophic hormone are important components of the melanocortin system. Melanocortin receptors (MCR, MC1R-MC5R) are important members of the G protein-coupled receptor superfamily. The latest clinical research data indicate that the expression of MC3R

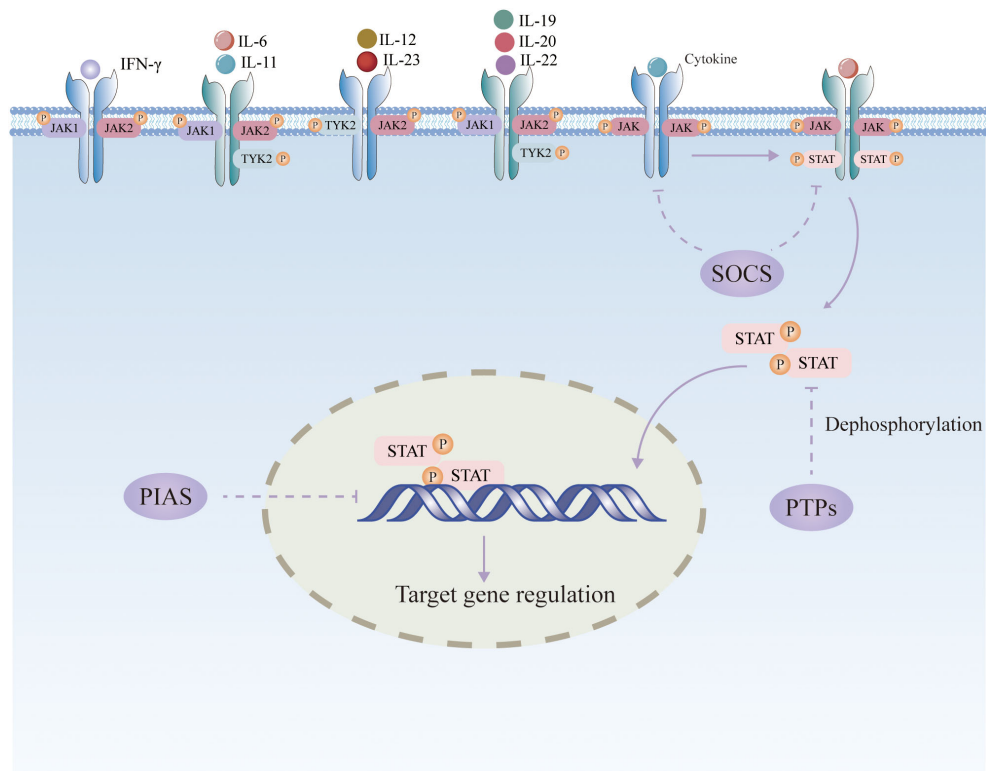


FIGURE 1
Schematic diagram of the JAK/STAT signaling pathway.

and MC5R is significantly increased in inflamed mucosa of inflammatory bowel disease (IBD) patients compared to normal mucosa (17). Importantly, the melanocortin system plays a key role in inflammation and immune regulation (18, 19). The melanocortin system is involved in the development of IBD through multiple pathways (20). Melanocortin peptides, especially α -MSH, have potent anti-inflammatory and immunomodulatory activities (21). It has been suggested that α -MSH may indirectly affect the activity of the JAK/STAT signaling pathway by regulating cytokine production (22, 23). Melanocortin attenuates myocardial ischemia/reperfusion injury by activating JAK/STAT signaling (24). Seemingly paradoxically, α -MSH was shown to activate the JAK2/STAT1 pathway by binding its MC5R receptor (25). The regulatory mechanism of melanocortin system on the JAK/STAT pathway remains to be further investigated in depth.

2.4 Cross-talk between the JAK/STAT pathway and other signaling networks

Diverse components of the JAK/STAT pathway, such as JAK, STAT, receptors, and gene transcription factors, are embedded in a dynamic cross-talk with other signaling networks. For example, the cross-talk between nuclear factor-kappa-B (NF- κ B) and STAT3 has been observed in numerous inflammatory disorders and cancers. First, IL-6, a gene production regulated by NF- κ B pathway, serves as a critical STAT3 activator (26). Second, STAT3-mediated

acetylation of NF- κ B p65 enhances its transcriptional activity in the nucleus and promotes the expression of pro-inflammatory factors such as IL-6 and TNF- α (27). Finally, STAT3 stimulates the expression of p52 and CD30, which induces sustained activation of non-canonical NF- κ B signaling (28). Furthermore, dimerization of IL-6-type cytokine receptors not only activates the JAK/STAT signaling pathway, but also induces the mitogen-activated protein kinase (MAPK) cascade by recruiting SH2-domain-containing tyrosine phosphatase (SHP2) to tyrosine-phosphorylated gp130 and phosphorylating it in a JAK1-dependent manner. The phosphorylated SHP2 combines with the growth factor receptor-bound protein/Son of Sevenless (Grb2-SOS) complex, resulting in the activation of the Ras-Raf-MAPK pathway (29). Autophagy in epithelial cells is usually considered to exert a protective effect in UC (30). On the one hand, it was found that the activation of JAK2/STAT3 pathway directly suppressed the transcription of autophagy regulator Beclin-1, contributing to the inhibition of autophagy and the initiation of intestinal cell death (31). On the other hand, autophagy promotes IFN- γ -induced Jak2/STAT1 activation by inhibiting the expression of reactive oxygen species and SHP2 (32).

Other inflammatory pathways such as the NF- κ B pathway also play important roles in inflammation, but the NF- κ B pathway is widely involved in a variety of physiopathologic processes with relatively limited specificity. In contrast, the JAK/STAT pathway is more direct and critical in cytokine-mediated inflammatory signaling. Thus, targeting the JAK/STAT pathway enables more precise intervention in the inflammatory process of UC. Besides, the

JAK/STAT pathway involves diverse cytokines and immune cells. Compared to this, the regulatory scope of NF- κ B is relatively limited. Natural products may target the JAK/STAT pathway to comprehensively regulate the inflammatory response through multi-target effects. Importantly, JAK inhibitors (e.g., tofacitinib) have shown significant efficacy in the treatment of UC, further validating the feasibility of the JAK/STAT pathway as a therapeutic target (33). In addition, some inflammatory pathways have complex regulatory mechanisms, making intervention difficult. For example, there are multiple upstream and downstream kinases in the MAPK pathway, with complex interactions between members. Comprehensive intervention may induce more adverse effects. The JAK/STAT pathway is comparatively clear and its intervention by natural products has been more intensively studied. Consequently, JAK/STAT a privileged target for natural products in UC.

3 Role of JAK/STAT pathway in the pathogenesis of UC

3.1 Influence on inflammatory response

The JAK2/STAT3 axis is a major pathway for transcription factors associated with mediating proinflammatory cytokine in intestinal mucosal inflammation. Inflammatory factors such as interferon- γ (IFN- γ) and interleukin (IL) have been found to promote the activation of JAK/STAT pathway, which in turn

exerts immunomodulatory functions (34, 35). In recent years, increasing evidence suggested that aberrant activation of the JAK/STAT signaling pathway is related to the pathogenesis of UC. The expression of four JAK genes was upregulated in the intestinal mucosal epithelium of patients with active UC (36). Polymorphisms in JAK2 and STAT3 genes correlate with the severity of UC patients (37). Most cytokines mediate inflammatory responses by activating JAK/STAT pathway in UC (summarized in Figure 2). Previous studies have revealed that IL-6 is involved in the pathogenesis of UC (38). IL-6 binding to its receptor activates JAK1/2 and TYK2 and contributes to the phosphorylation and transcriptional activation of STAT3, which ultimately regulates T cell differentiation and inflammatory response (35, 39). Moreover, IL-12 and IL-23 activate STAT3 and STAT4 through JAK2 and TYK2, respectively (40, 41).

3.2 Impact on intestinal epithelial cells

Intestinal mucosal structures are maintained by a balance between apoptosis and proliferation of intestinal epithelial cells (IECs). However, IECs in UC patients exhibit a higher rate of apoptosis (42). Abnormal apoptosis leads to the loss of intestinal epithelial structures, disrupting the intestinal mucosal barrier and further activating excessive immune responses, eventually leading to uncontrollable inflammatory responses and mucosal damage. The JAK pathway is known to play an influential role in the regulation of cell proliferation and apoptosis (43). Studies showed

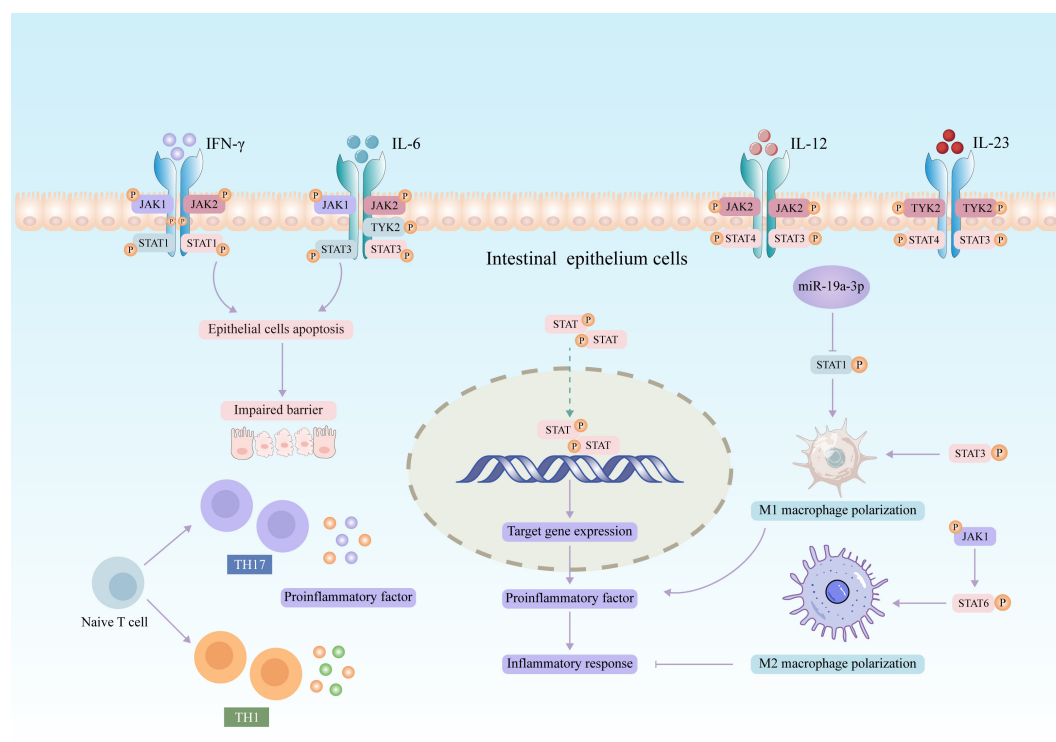


FIGURE 2
Potential role of JAK/STAT signaling pathway in the pathogenesis of UC.

that the activation of STAT1 or STAT3 promotes apoptosis in IECs (44, 45). miR-124-3p can directly target the STAT3 3'-UTR to modulate STAT3 expression (46). A recent study indicated that the overexpression of miR-124-3p attenuates apoptosis and reactive oxygen species production by targeting STAT3 in lipopolysaccharide (LPS)-induced colonocytes (47).

3.3 Modulation of intestinal macrophages

Under homeostatic conditions, circulating lymphocyte antigen 6 complex (Ly6C) monocytes in mice and CD14 monocytes in humans are constantly attracted to the intestinal tract and differentiate into mature F4/80 macrophages with high levels of CX3C chemokine receptor 1 (CX3CR1^{hi}). CX3CR1 macrophages directly activate neighboring T cells to amplify the inflammatory response (48). A recent study found that gut-resident CX3CR1^{hi} macrophages trigger tertiary lymphoid structures and IgA response *in situ* (49). Furthermore, intestinal mucosal CXCR4⁺ IgG plasma cells drive the activation of CD4 macrophage and exacerbate UC (50). Compared to the lamina propria of the normal mucosa, the number of macrophages is greatly increased and activated in the localized colonic tissues of individuals with active UC, indicating that intestinal macrophages are implicated in the occurrence and progression of UC (51). Particularly, different expression of Tim-4 and CD4 can divide intestinal macrophages into three subsets, including locally maintained macrophages (Tim-4⁺CD4⁺), circulating monocyte-renewing macrophages (Tim-4⁻CD4⁺), and macrophages with the high monocyte-replenishment rate (Tim-4⁻CD4⁻) (52). Furthermore, macrophages from colonic lamina propria cells can be divided into subpopulations based on the expression of F4/80 and CD11b. F4/80^{hi} macrophages are considered to be intestinal resident macrophages, whereas CD11b^{hi} macrophages are regarded as infiltrative macrophages supplemented by circulating monocytes (53, 54). Interestingly, JAK/STAT signaling has an important regulatory effect on macrophage (55–58). It was found that inhibition of the JAK2/STAT3 pathway resulted in a significant reduction in apoptosis, collagen deposition, and immunoreactivity of intestinal macrophages (59). Notably, the levels of IFN- γ are markedly elevated in the mucosa of IBD patients. This cytokine promotes the pro-inflammatory characteristics of CD14^{hi} macrophages in humans (60). Consistently, the complete deletion of IFN γ R1 or its downstream transcription factor STAT1 suppresses the formation of immature Ly6C MHCII macrophages (61).

Macrophages are highly plastic in different environments, exhibiting different phenotypes and functions depending on microenvironmental stimuli and signals (62). Macrophages are divided into classically activated M1-type macrophages with proinflammatory effects and alternatively activated M2-type macrophages with anti-inflammatory effects (63, 64), both of which are involved in UC pathology (65). Recently, a growing amount of studies indicated that STAT1 plays a critical role in the modulation of M1 macrophage polarization (66–68). It was reported that miR-19a-3p inhibited M1 macrophage polarization as an upstream regulator of STAT1 (69). Moreover, SOCS3-

deficient macrophages showed increased STAT3 expression and M1 polarization (70). Interestingly, the JAK1/STAT6 pathway is an important pathway in the induction of M2 macrophage polarization (71). IL-4 binds to its receptor to activate JAK1, which recruits STAT6 phosphorylation and promotes the expression of M2 macrophage markers (72).

Notably, most of the current studies have been conducted mainly using *in vitro* bone marrow-derived macrophages (BMDMs). When cultured *in vitro*, BMDMs rely on specific cytokines to induce differentiation. However, the induction process is significantly different from the complex intestinal microenvironment *in vivo*. Moreover, BMDMs differ from intestinal macrophages in their degree of differentiation and maturation, leading to their distinct performance in the expression and activity of some key functional proteins. Apart from participating in immune defense, intestinal macrophages also play a crucial role in maintaining intestinal homeostasis and regulating intestinal microbiota balance. On the other hand, BMDMs lack gut-related signaling stimulation in the *in vitro* environment to fully exhibit these complex functions of intestinal macrophages.

3.4 Regulation of T cell balance

T cells are another important type of immune cells involved in adaptive immunity. Studies demonstrated that the JAK/STAT signaling pathway is critical in modulating T cell differentiation (35, 73, 74). A recent study showed a significant increase in the number of CD4 T cells in UC patients (75). Naive CD4 T cells are induced to differentiate into different types of T cells in different cytokine microenvironments, including T helper cell 1 (Th1), Th2, Th17, and regulatory T cells (Tregs). Abnormally activated CD4 T cells differentiate into subpopulations of Th1 and/or Th17 cells that subsequently infiltrate the colon to mediate autoimmune responses in UC (76). Cytokines such as Th1-induced IL-2 and IFN- γ , and Th17-induced IL-17 and IL-21 promote inflammatory responses and exacerbate colitis (77). In contrast, Tregs control effector T cell immunosuppression through intercellular contacts or secretion of anti-inflammatory cytokines. Th17/Treg balance facilitates the maintenance of intestinal immune homeostasis, an imbalance of which is the source of immune dysfunction in intestinal mucosa (78). Increasing data suggested that proinflammatory cytokines that stimulate the JAK/STAT signaling pathway govern the differentiation of naive Th1 and Th17 cell subsets and aggravate the development of UC (35, 73). STAT5 and forkhead box P3 (Foxp3) are key transcription factors for Tregs, whereas retinoic acid-related orphan receptor γ (ROR γ) and STAT3 are key transcription factors for Th17 cells (79, 80). The overactivation of STAT3 promotes the Th17-like transformation of Treg and exacerbates immune responses (81). IL-12 or IFN- γ binds to their receptors to activate STAT1, STAT4, and the T-box transcription factor, driving the differentiation and function of Th1 cells. Similarly, IL-6 binds to its receptor and drives Th17 differentiation by activating ROR γ and STAT3 (82, 83). Interestingly, TAK-242, a specific inhibitor of Toll-like receptor-4

(TLR4), was shown to alleviate UC by regulating macrophage polarization and Th homeostasis through the TLR4/JAK2/STAT3 signaling pathway (84).

4 Natural products involved in the regulation of JAK/STAT signaling in UC

4.1 Glycosides

Arbutin (molecular formula: $C_{12}H_{16}O_7$, molecular weight: 272.25) is a glycoside compound mainly extracted from the leaves of arbutus. The chemical structure of arbutin is shown in Figure 3. Arbutin is a hydroquinone glucoside, with two different configurations: α and β arbutin. Compared to α -arbutin, β -arbutin is more frequently found in nature and typically occurs in higher concentrations in plants. β -arbutin has been widely researched for its whitening, anti-inflammatory, antimicrobial, antioxidant, and anticancer properties (85). Arbutin has been reported to significantly down-regulate the levels of inflammatory cytokines (IL-1 β , IL-6, and TNF- α), iNOS, and cyclooxygenase-2 (COX-2) in

colitis mice (86). In addition, arbutin remarkably inhibited the phosphorylation of JAK2 and STAT3 and suppressed IECs apoptosis, thereby improving barrier function (Table 1). *In vivo* experiment demonstrated that p-JAK2 expression was significantly inhibited by arbutin and AG490, a JAK2 inhibitor (86). No additional therapeutic efficacy was observed with the combination of arbutin and AG490. *In vitro* experiment showed that the inhibitory effect of arbutin on p-STAT3 and inflammatory factors (TNF- α and IL-6) was significantly reversed by AG490, further suggesting that arbutin may be a potential JAK2 inhibitor. These results indicated that the effect of arbutin on JAK was primary rather than secondary to broader anti-inflammatory effects. Interestingly, a recent study found that arbutin also inhibited the formation of neutrophil extracellular traps and increased the diversity and abundance of gut microbiota (87).

4.2 Polysaccharides

Chrysanthemum morifolium Ramat (Juhua), as a medicinal and edible homeopathic plant with strong heat-removing and detoxifying abilities, has long been widely used for the treatment of various diseases, including influenza, colitis, stomatitis,

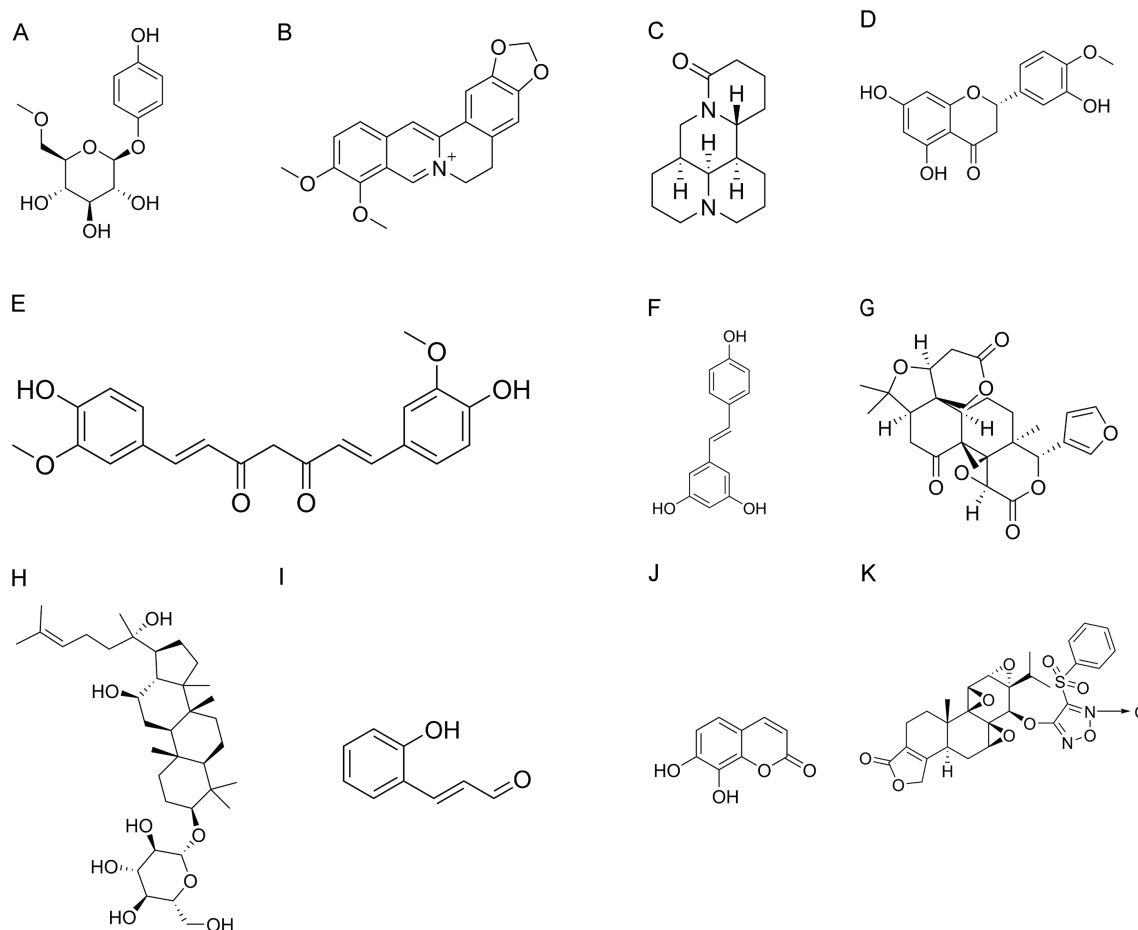


FIGURE 3

Chemical structures of natural products. (A) Arbutin. (B) Berberine. (C) Matrine. (D) Hesperetin. (E) Curcumin. (F) Resveratrol. (G) Limonin. (H) Ginsenoside Rh2. (I) 2'-Hydroxycinnamaldehyde. (J) Daphnetin. (K) ZT01.

TABLE 1 Summary of natural compounds involved in the regulation of JAK/STAT signaling in UC.

Phytochemicals	Type	Experimental model	Mechanisms	References
Arbutin	Glycoside	DSS-induced UC mice, LPS-stimulated IEC-6 and RAW264.7 cells	↓TNF- α , IL-1 β , and IL-6, ↓iNOS and COX2, ↑Bcl2, ↓MLCK, ↓p-JAK2, p-STAT3, and SOCS3	(86)
<i>Chrysanthemum</i> polysaccharide	Polysaccharide	TNBS-induced colitis rats	↓TNF- α , IFN- γ , IL-6, and IL-1 β , ↓MDA and MPO, ↑SOD, ↓p-p65, TLR4, p-STAT3, and p-JAK2	(94)
Tetrastigma hemsleyanum polysaccharide	Polysaccharide	DSS-induced UC mice, Caco-2 cells induced by LPS in combination with IL-6	↑Claudin-1, ↓IL-6, TNF- α , MCP-1, and IFN- γ , ↓IL-17A, ↑Foxp3, ↓p-JAK2, JAK2, p-STAT3, and STAT3, ↑SOCS1	(97)
Aloe polysaccharide	Polysaccharide	TNBS-induced colitis rats, HT-29 cells stimulated by TNF- α and LPS	↓IL-6, ↓p-JAK2, JAK2, p-STAT3, and STAT3	(7)
Berberine	Alkaloid	DSS-induced UC mice	↓NLRP3, ASC, and cleaved caspase-1, ↓IL-1 β , IL-6, and IL-18, ↑ZO-1, E-cadherin, occludin, claudin-1, and MUC2, ↓OSM and OSMR, ↓p-JAK1, p-JAK2, p-STAT1, and p-STAT3	(112)
Matrine	Alkaloid	DSS-induced NCM460 cells	↓TNF- α , IL-1 β , IL-2, and IL-6, ↓MPO and NO, ↓Bax and cleaved caspase-3, ↑Bcl-2, ↓p-JAK2/JAK2 and p-STAT3/STAT3	(116)
Hesperetin	Flavonoid	TNBS-induced colitis rats	↑GSH and SOD, ↓NO content, ↓IL-6, NF- κ B, RAGE, and TNF- α , ↓p-JAK2 and p-STAT3, ↑SOCS3	(117)
Curcumin	Polyphenol	DSS-induced UC mice	↓IL-7, IL-15, and IL-21, ↓JAK1, STAT5, and p-STAT5, ↑PIAS1	(122)
			↓p-STAT3, ↓DNA-binding activity of STAT3 dimers, ↓MPO, IL-1 β , and TNF- α	(123)
		TNBS-induced colitis mice	↓p-JAK2, p-STAT3, and p-STAT6, ↑SOCS1, SOCS3, and PIAS3, ↓Activation of dendritic cells	(119)
Resveratrol	Polyphenol	DSS-induced UC mice, HCT116 cells	↓IL-6, IL-1 β , and TNF- α , ↑IL-10, ↓O-GlcNAcylation, ↓p-JAK2 and p-STAT3, ↓NOS2 and COX2	(133)
Ginsenoside Rh2	Terpenoid	DSS-induced UC mice, IL-6-stimulated NCM460 cells	↓TNF- α , IL-6 and IL-1 β , ↓p-STAT3, ↓miR-214, ↑PTEN	(138)
Limonin	Terpenoid	DSS-induced UC mice, IL-6-stimulated NCM460 cells	↓IL-6 and TNF- α , ↑IL-10, p-STAT3, ↓miR-214, ↑PTEN and PDLIM2	(140)
2'-Hydroxycinnamaldehyde	Other types	DSS-induced UC mice, LPS-treated FHC cells	↓TNF- α , IL-6 and IL-1 β , ↑IL-10, ↑ZO-1, occludin, Bcl-2, E-cadherin, and Claudin-3, ↓Bax, ↓p-STAT3 and the translocation of STAT3 from cytoplasm to nucleus	(145)
Daphnetin	Other types	DSS-induced mice, LPS-challenged Caco-2 cells	↓TNF- α , IFN- γ , IL-6, and IL-1 β , ↑ZO-1, occludin, and BCL-2, ↓Bax and cleaved caspase 3, ↓MDA and SOD, ↓JAK2 and STAT3	(152)
ZT01	Other types	DSS-induced UC mice, BMDMs stimulated by LPS and IFN- γ	↑ZO-1 and E-cadherin, ↓p-JAK1, p-JAK2, p-STAT1, and p-STAT3, ↓the differentiation of Th1 and Th17 cell, ↓the proinflammatory macrophage phenotype polarization	(154)

The symbol “↓” indicates inhibition, while “↑” represents promotion.

cardiovascular diseases, and various chronic diseases (88). Polysaccharide is one of the key active components in *Chrysanthemum morifolium* Ramat. It exhibits anti-inflammatory, antioxidant, immunomodulatory, anti-cancer, hepatoprotective, and gastrointestinal function regulatory effects (89, 90). The monosaccharides of *Chrysanthemum* polysaccharides (CP) mainly include galactose, glucose, arabinose, and mannose (89). Several studies have reported that the biological activity of CP is related to their chemical properties, molecular weights, and monosaccharide composition (91, 92). The high antioxidant activity of snow CP is partly attributed to the low molecular weight and high content of unmethylated galacturonic acid (93). In rats with 2, 4, 6-trinitrobenzene sulfonic acid (TNBS)-induced colitis, CP reduced the expression of proinflammatory cytokines and blocked the phosphorylation of STAT3 and JAK2, resulting in significant relief of colitis (94). Furthermore, CP influenced biomarkers and metabolic pathways in plasma and urine. Although this study found that CP decreased the expression of IL-6/JAK2/STAT3 pathway-related proteins and mRNAs, it was not clear whether the effect of CP on JAK/STAT was primary or secondary to broader anti-inflammatory actions. For a deeper understanding of its mechanism of action, further experiments need to be designed to distinguish the primary and secondary effects. For example, after treatment with specific pathway inhibitors or activators, the changes in the effects of CP on the JAK/STAT pathway and inflammatory response should be observed.

Tetrastigma hemsleyanum Diels et Gilg (Sanyeqing) is a traditional herb native to China. Its whole plant is medicinally used and it is of great concern for its remarkable medicinal value. In particular, it shows strong potential in anti-inflammatory, antibacterial, and antiviral properties (95). *Tetrastigma hemsleyanum* Diels et Gilg is rich in active substances such as flavonoids, phenolic acids and polysaccharides. *Tetrastigma hemsleyanum* polysaccharide (THP) consists of various monosaccharides, mainly including galactose, glucuronic acid, mannose, glucose, rhamnose, and arabinose (96). THP has shown great potential in the treatment of UC (97, 98). THP reduces the expression of the proinflammatory cytokines IL-6, TNF- α , and IL-17 and promotes the regulatory factors forkhead box protein P3 and Tregs (97). Importantly, it exerts anti-inflammatory effects by promoting SOCS1 expression and inhibiting JAK2/STAT3 signaling (97). Additionally, THP elevates levels of tight junction proteins in colonic tissues and decreases colonic permeability, improving the intestinal mucosal barrier. Interestingly, it can also modulate the gut microbiota structure and corresponding short-chain fatty acid metabolites in mice with IBD (98). Notably, although most studies used p-JAK2 and p-STAT3 as efficacy biomarkers, they did not distinguish whether natural products inhibited them directly or regulated them indirectly through upstream cytokines or microbiota-derived metabolites. The specific mechanism by which natural products regulate JAK/STAT signaling is currently not precise enough. Follow-up studies could construct STAT knockdown models by CRISPR/Cas9 technology to verify the specific action targets of active ingredients.

Moreover, relevant upstream regulators can be knocked down or overexpressed to further explore the regulation mechanism of STAT3 phosphorylation by THP.

Aloe polysaccharide is an active macromolecule extracted from *Aloe vera*. It shows excellent protection against ulcers and significantly prevents ulcer formation (99). *In vivo* and *in vitro* experiments have demonstrated that aloe polysaccharide effectively ameliorated apoptosis in colon tissue by inhibiting the JAK2/STAT3 signaling pathway (7). Meanwhile, aloe polysaccharide contributed to the reduction of IL-6 levels and restoration of colon length in TNBS-induced UC rats. Similarly, this study only used methods such as immunohistochemistry, Western blot, and RT-PCR to detect the expression level of the JAK/STAT pathway, with insufficient depth and specificity of the mechanism of action.

4.3 Alkaloids

Berberine (molecular formula: $C_{20}H_{18}NO_4^+$, molecular weight: 336.4), a natural pentacyclic isoquinoline alkaloid, is the most representative and abundant constituent of the TCM *Coptis chinensis* Franch (Huanglian). Berberine contains two benzene rings, an isoquinoline ring, and functional groups such as methoxy groups, which endow it with unique biological activities. In addition to its anti-inflammatory and antioxidant activities, berberine exhibits a variety of pharmacological effects, including anti-apoptotic, antitumor, hepatoprotective, and cardiovascular protective effects (100–102). Interestingly, it is widely known for its anti-inflammatory effects in inflammatory gastrointestinal diseases (103). In a double-blind phase I trial, berberine was demonstrated to improve colonic mucosal histologic scores in Chinese patients with UC (104). Furthermore, the Xijing Hospital of Digestive Diseases is currently undertaking a phase IV clinical trial to determine the impact of berberine on the annual recurrence rate of UC in remission (NCT02962245, ClinicalTrials.gov). Interestingly, a recent *ex vivo* study explored the synergistic effects of berberine, *Hericium erinaceus*, and quercetin, providing a more effective therapeutic option for UC patients. Their combination reduced the expression of proinflammatory cytokines and promoted the expression of the anti-inflammatory cytokine IL-10 in IBD tissues (105). Berberine has been discovered to relieve experimental colitis by altering the inflammatory response of immunological and epithelial cells, improving intestinal barrier function, and modulating intestinal microbiota (106–109). Oncostatin M (OSM) belongs to the IL-6 cytokine family and is primarily produced by activated macrophages, neutrophils, dendritic cells, and T cells (110). Previous studies have confirmed that recombinant OSM induces the activation of the JAK-STAT pathway via a heterodimeric receptor consisting of OSMR and gp130 (111). Importantly, berberine has been found to alleviate intestinal fibrosis by inhibiting the OSM-mediated JAK-STAT pathway and interfering with the interaction between intestinal stromal cells and immune cells (112). Furthermore, berberine was shown to inhibit M1 macrophage polarization and induce M2 macrophage polarization, by activating the IL-4-STAT6 signaling pathway, thereby exerting a therapeutic effect on UC (113).

Radix Sophorae Flavescentis is the dried root of *Sophora flavescens* Aiton (Kushen), belonging to the Leguminosae family. It is a promising traditional herb with the effect of clearing heat and dampness and has long been used to treat UC. Alkaloids and flavonoids are the main components of *Sophora flavescens* Aiton. Matrine is isolated from the roots of *Sophora flavescens* Aiton, *Sophora tonkinensis*, and *Sophora alopecuroides* (Kudouzi). Matrine is a tetracyclic quinolizidine alkaloid with the chemical formula $C_{15}H_{24}N_2O$ and a molecular weight of 248.36. Matrine exhibits a wide range of pharmacological activities, including analgesic, anticancer, anti-inflammatory, antiviral, antifibrotic, and immunomodulatory effects (114). Because of its anti-inflammatory and immunomodulatory properties, matrine has great potential in the treatment of UC (115). Apart from this, matrine improved the composition and function of intestinal microbiota in mice with dextran sulfate sodium (DSS)-induced colitis. It decreased the proportions of *Firmicutes*, *Bacteroidetes*, and *Proteobacteria*, increasing the relative abundance of *Lactobacillus* and *Akkermansia* (115). A recent study confirmed that matrine inhibited proinflammatory factors, MPO activity, NO production, and apoptosis, thus effectively alleviating UC (116). Furthermore, matrine was found to suppress the phosphorylation levels of JAK2 and STAT3, but did not affect the phosphorylation of STAT5.

4.4 Flavonoids

Hesperetin (molecular formula: $C_{16}H_{14}O_6$, molecular weight: 302.28) is a naturally occurring flavonoid compound in citrus fruits and is widely found in various traditional herbal medicines such as grapefruit peel, orange peel, and tangerine peel. In TNBS-induced UC rats, hesperetin significantly enhanced glutathione levels and superoxide dismutase activity to reduce colonic oxidative stress, while significantly reducing NO levels (117). Hesperetin also mitigated the inflammatory injury by significantly decreasing IL-6 as well as inhibiting the expression of NF- κ B, receptor for advanced glycation end products, and TNF- α . In addition, hesperetin significantly inhibited the phosphorylation of JAK2 and STAT3 and promoted the expression of SOCS3, thereby alleviating colitis. As mentioned above, the present study did not clarify whether natural products directly inhibit JAK2/STAT3 phosphorylation through small-molecule binding or indirectly modulate this pathway through upstream cytokines. Natural products may affect JAK2 and STAT3 phosphorylation through different mechanisms in different studies. If all do not distinguish between direct and indirect regulation and only use them as biomarkers of efficacy, it will cause incomparability between the results of studies.

4.5 Polyphenols

Curcumin (molecular formula: $C_{21}H_{20}O_6$, molecular weight: 368.4) is an active polyphenol obtained from the dry rhizomes of herbs such as turmeric and tulip. It is also considered one of the potential drugs for the treatment of UC (118). Curcumin could alleviate UC by inhibiting dendritic cell-mediated expression of

proinflammatory factors (119), modulating Th17/Treg homeostasis (120), and regulating M1/M2 macrophage polarization (121). Interestingly, the regulation of memory T cell homeostasis by curcumin is associated with the inhibition of JAK1/STAT5 signaling activity (122). In addition, curcumin not only suppressed STAT3 phosphorylation and STAT3 dimer binding to DNA, but also significantly inhibited the expression of proinflammatory cytokines, consequently ameliorating UC (123). Another study revealed that curcumin inhibited the phosphorylation of JAK2, STAT3, and STAT6 and upregulated the expression of downstream proteins (SOCS1, SOCS3, and PIAS3) in TNBS-induced UC rats (119). Moreover, curcumin inhibits dendritic cell activation and restores immune homeostasis by modulating the JAK/STAT/SOCS signaling pathway, effectively treating colitis (119). A randomized, double-blind, placebo-controlled trial demonstrated that the herbal combination of curcumin-QingDai significantly reduced the Disease Activity Index (DAI) score in patients with active UC and effectively induced their response and remission (CLINICALTRIALS: gov ID: NCT03720002).

Resveratrol (3,5,4'-trihydroxy-trans-stilbene) is a polyphenolic stilbenoid isolated from *Veratrum grandiflorum* and abundantly found in grapes, mulberries, peanuts, rhubarb and several other plants. It is a well-known antioxidant (124). Due to its planar stilbene motif, resveratrol exhibits relatively high hydrophobicity. As a result, it demonstrates a comparatively strong affinity for hydrophobic pockets and binding sites within proteins. Furthermore, the polar hydroxyl (OH) groups serve as both hydrogen-bond donors and acceptors. These groups are capable of establishing numerous interactions with amino acid side chains and backbone amide groups (125). Clinical and preclinical studies have demonstrated that resveratrol exerts protective effects in numerous disease models, including digestive diseases, cardiovascular diseases, diabetes, tumors, and neurodegenerative diseases, which may be related to its multi-targeting properties (126–129). Notably, resveratrol has been demonstrated to restore the homogeneity and diversity of gut microbiota to some extent in colitis mice (130). Moreover, dietary resveratrol attenuated the inflammatory status and down-regulated the expression of proinflammatory cytokines such as IL-2, IFN- γ , IL-1 β , IL-6, and TNF- α in colitis mouse model (131). Among the known resveratrol targets, JAK-STAT signaling has received widespread attention (132). It was shown that increased O-linked N-acetylglucosamine modification (O-GlcNAcylation) of STAT3 upregulated the expression of proinflammatory cytokines such as IL-6, IL-1 β , and TNF- α , while downregulating the level of the anti-inflammatory cytokine IL-10 and aggravating colitis in mice (133). In addition, the levels of COX-2 and iNOS were elevated. Encouragingly, resveratrol inhibited the O-GlcNAcylation of STAT3, thereby inhibiting its phosphorylation as well as the activity of JAK2/STAT3 pathway, and consequently alleviating IBD (133). Moreover, resveratrol induced Tregs in mice with colitis, which was dependent on the downregulation of miR-31 (134). Meanwhile, it suppressed inflammatory T cells (Th1 and Th17). A randomized, double-blind, placebo-controlled study showed that supplementation with 500 mg resveratrol for 6 weeks improved the quality of life and

reduced colonic inflammation in UC patients (135). Unfortunately, resveratrol's low bioavailability and poor water solubility restrict its therapeutic use. The stability and oral bioavailability of resveratrol should be improved by future research using different delivery methods and changes (136).

4.6 Terpenoids

Ginseng, a traditional herbal medicine, is the dried root of *Panax ginseng* C. A. Meyer., a plant of the family Wujiaceae. As a valuable medicinal herb, it has been used in China for more than 2,000 years. Ginsenoside Rh2 (molecular formula: $C_{36}H_{62}O_8$, molecular weight: 622.9) is one of the active ingredients extracted from ginseng root. It possesses various pharmacological activities and has great potential in the treatment of UC (137). *In vivo* and *in vitro* experiments revealed that ginsenoside Rh2 effectively inhibited STAT3 phosphorylation and miR-214 expression (138). Ginsenoside Rh2 was found to indirectly suppress STAT3 phosphorylation by inhibiting the upstream cytokine IL-6.

Limonin (molecular formula: $C_{26}H_{30}O_8$, molecular weight: 470.5) is a triterpenoid derived from citrus and possesses favorable anti-inflammatory and antiapoptotic effects. Limonin reduced the generation of proinflammatory cytokines TNF- α , IL-1 β , and IL-6 as well as the expression of inflammatory proteins COX-2 and iNOS in the colonic tissues of mice with DSS-induced colitis (139). Moreover, limonin was found to ameliorate DSS-induced chronic colitis in mice by inhibiting the endoplasmic reticulum-stressed PERK-ATF4-CHOP pathway and NF- κ B signaling (139). In addition to this, limonin also improved the prognosis of UC by downregulating p-STAT3/miR-214 levels (140).

4.7 Other types

2'-Hydroxycinnamaldehyde (HCA) (molecular formula: $C_9H_8O_2$, molecular weight: 148.16) is an active component isolated from the stem bark of *Cinnamomum cassia* (Rougui) (141). HCA was proved to have anticancer, anti-inflammatory, antioxidant, and immunomodulatory effects (141–143). Interesting, HCA was screened as a natural STAT3 inhibitor (141, 144). A recent study showed that HCA directly binds to STAT3 and inhibits its activation (145). The hydroxyl group of HCA may interact with the protein-binding site of STAT3 via hydrogen bonding to enhance binding specificity. Thanks to this property, it inhibits inflammatory cytokine expression, reduces apoptosis of IECs, and attenuates intestinal mucosal barrier damage, thus effectively alleviating UC (145).

Daphnetin (molecular formula: $C_9H_6O_4$, molecular weight: 178.14), a coumarin derivative isolated from the Daphne plant, is a natural compound with multiple therapeutic potential (146–148). Daphnetin possesses oxygen-containing heterocycles with a characteristic benzo- α -pyrone framework (149). The catechol moiety served as the crucial pharmacophore for the antioxidant activity of daphnetin (149). Apart from its antioxidant activity,

daphnetin also exhibits diverse therapeutic potentials, including anti-inflammatory, analgesic, antibacterial, neuroprotective, hepatoprotective, nephroprotective, and anticancer activities (150). A previous study has demonstrated that daphnetin ameliorates colitis by regulating microbiota composition and TH17/Treg balance (151). A recent study reported that Daphnetin attenuated intestinal inflammation, oxidative stress, and apoptosis in UC, which was associated with the inhibition of REG3A-dependent JAK2/STAT3 signaling (152).

Triptolide, a natural diterpenetriepoxide which is isolated from *Tripterygium wilfordii* Hook F (Leigongteng), has prominent anti-inflammatory and immunosuppressive properties. ZT01 is a newly obtained tretinoin derivative with strong anti-inflammatory effects and low toxicity (153). ZT01 may be an attractive candidate for future development as an anti-UC drug. Importantly, ZT01 significantly inhibits T cell differentiation into Th1 or Th17 cell subsets and prevents macrophage polarization to an inflammatory phenotype by modulating the JAK/STAT signaling pathway (154).

5 Conclusion and perspective

The increasing incidence of UC has placed a heavy burden on the global health system. The JAK/STAT signaling pathway mediates the pathogenesis of UC to some extent. There are limitations to some of the current studies on the JAK/STAT pathway and UC. The sample sizes of the studies detecting the activation level of the JAK/STAT pathway in colon biopsies from UC patients are small. Although it is possible to obtain information on a specific patient group to some extent, it is difficult to fully reflect the real situation of the entire UC patient population and is prone to bias. Moreover, single-center studies may be affected by factors such as geography and medical level, which makes the generalizability of the findings questionable. Future studies need to expand the sample size, use multicenter studies, and deeply investigate the reasons for the differences in JAK/STAT pathway activation. In terms of findings, higher levels of JAK and STAT expression were detected in the inflamed colonic mucosa of UC patients compared to the uninflamed mucosa. However, the current study did not further investigate the reasons for this difference, whether it is genetic differences in individuals, living environment or other factors. The lack of in-depth analysis would limit a comprehensive understanding of the pathogenesis of UC.

In recent years, natural products have received extensive attention from the medical community. They have the advantages of multiple pathways and multiple targets. It has been demonstrated that herbal active ingredients alleviate UC through various pathways, such as targeting the JAK/STAT pathway to reduce intestinal inflammation, improving the function of IECs, regulating Th17/Treg balance, and modulating macrophage status. This review systematically summarized the recent advances in natural products targeting the JAK/STAT pathway to treat UC, including polysaccharides, alkaloids, polyphenols, terpenoids, flavonoids, glycosides, and other types of compounds. Natural products are potential candidates to treat UC by targeting the JAK/STAT pathway.

Nevertheless, there are many challenges. Firstly, the current studies on the targeting of the JAK/STAT pathway by natural products for the treatment of UC mainly focus on the animal and cellular experimental level, which cannot be fully equated with the immunohistopathology of UC patients. At present, there is insufficient research on the pharmacokinetic properties of many natural products, such as their absorption, distribution, metabolism, and excretion in the body. These uncertainties affect the design of standardized dosages and the formulation of dosing regimens. Furthermore, natural products such as resveratrol have poor water solubility and low bioavailability, making it difficult to make suitable dosage forms for clinical use. There is an urgent need to improve the stability, solubility, and bioavailability of natural products to overcome the transformation challenges. Besides, when herbal active ingredients are combined with other drugs, they may affect pharmacokinetics and pharmacodynamics through multiple pathways, but their specific mechanisms and links are difficult to be clearly defined. These limitations pose a great challenge to mechanism research and efficacy assessment in the translation process of TCM. Preclinical and clinical studies are needed to validate the safety and efficacy of herbal active ingredients for the treatment of UC. Secondly, crosstalk exists between the JAK/STAT pathway and other signaling pathways, which means that targeting only one of JAK or STAT may not be sufficient for significant therapeutic effects. Exactly how natural products interfere with the JAK/STAT pathway and whether they interact with other signaling has not been fully elucidated. Therefore, subsequent scholars still need to conduct profound research on the mechanism of herbal active ingredients in the treatment of UC. Finally, the vast majority of studies have been limited to the effect of natural products on the JAK/STAT pathway and have not analyzed the in-depth laws between their chemical structures and pharmacological activities. The structural features of phytochemicals may influence their specificity for JAK/STAT through factors such as molecular size, shape, functional groups, charge distribution, and conformational flexibility. Although the intrinsic laws have not been fully revealed, structure-activity relationship studies and molecular docking will provide important clues for understanding these interactions.

Author contributions

DL: Conceptualization, Investigation, Supervision, Visualization, Writing – original draft. CM: Conceptualization, Formal analysis, Investigation, Writing – original draft. WZ: Conceptualization, Investigation, Writing – original draft. YZ: Funding acquisition,

Supervision, Writing – review & editing. YX: Funding acquisition, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This study was funded by the National Natural Science Foundation of China (No. 82374426), the Postgraduate Scientific Research Innovation Project of Hunan Province (CX20240748), the Postgraduate Scientific Research Innovation Project of Hunan University of Chinese Medicine (2024CX022), the Domestic First-class Construction Discipline of Chinese Medicine in Hunan University of Chinese Medicine, and the Construction Project of Inheritance Studio of National Famous Traditional Chinese Medicine Experts of National Administration of Traditional Chinese Medicine ((2022) No. 75).

Acknowledgments

We thank Jun Xu for her assistance with language.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Generative AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Le Berre C, Honap S, Peyrin-Biroulet L. Ulcerative colitis. *Lancet*. (2023) 402:571–84. doi: 10.1016/S0140-6736(23)00966-2
2. Kirchgerner J, Lemaitre M, Carrat F, Zureik M, Carbonnel F, Dray-Spira R. Risk of serious and opportunistic infections associated with treatment of inflammatory bowel diseases. *Gastroenterology*. (2018) 155:337–46.e10. doi: 10.1053/j.gastro.2018.04.012
3. Xue C, Yao Q, Gu X, Shi Q, Yuan X, Chu Q, et al. Evolving cognition of the JAK-STAT signaling pathway: autoimmune disorders and cancer. *Signal Transduct Target Ther*. (2023) 8:204. doi: 10.1038/s41392-023-01468-7
4. Gravina AG, Pellegrino R, Esposito A, Cipullo M, Romeo M, Palladino G, et al. The JAK-STAT pathway as a therapeutic strategy in cancer patients with immune

checkpoint inhibitor-induced colitis: A narrative review. *Cancers (Basel)*. (2024) 16:611. doi: 10.3390/cancers16030611

5. Cordes F, Lenker E, Weinlage T, Spille LJ, Bettenworth D, Varga G, et al. Impaired IFN- γ -dependent STAT3 activation is associated with dysregulation of regulatory and inflammatory signaling in monocytes of ulcerative colitis patients. *Inflammation Bowel Dis*. (2021) 27:887–901. doi: 10.1093/ibd/izaa280
6. van Gennep S, Fung I, Jong DC, Ramkisoen RK, Clasquin E, de Jong J, et al. Histological outcomes and JAK-STAT signalling in ulcerative colitis patients treated with tofacitinib. *J Crohns Colitis*. (2024) 18:1283–91. doi: 10.1093/ecco-jcc/jjae031
7. Lin H, Honglang L, Weifeng L, Junmin C, Jiantao Y, Junjing G. The mechanism of alopolyasaccharide protecting ulcerative colitis. *BioMed Pharmacother*. (2017) 88:145–50. doi: 10.1016/j.biopha.2016.11.138
8. Sandborn WJ, Su C, Sands BE, D'Haens GR, Vermeire S, Schreiber S, et al. Tofacitinib as induction and maintenance therapy for ulcerative colitis. *N Engl J Med*. (2017) 376:1723–36. doi: 10.1056/NEJMoa1606910
9. Tahvilian N, Masoodi M, Faghihi Kashani A, Vafa M, Aryaeian N, Heydarian A, et al. Effects of saffron supplementation on oxidative/antioxidant status and severity of disease in ulcerative colitis patients: A randomized, double-blind, placebo-controlled study. *Phytother Res*. (2021) 35:946–53. doi: 10.1002/ptr.6848
10. Xiao S, Yan Y, Shao M, Zhou X, Niu Z, Wu Y, et al. Kujieling decoction regulates the Treg/Th17 cell balance in ulcerative colitis through the RA/RAR α signaling pathway. *J Ethnopharmacol*. (2024) 318:116909. doi: 10.1016/j.jep.2023.116909
11. Degreys S, de Bock CE, Cox L, Demeyer S, Gielen O, Mentens N, et al. JAK3 mutants transform hematopoietic cells through JAK1 activation, causing T-cell acute lymphoblastic leukemia in a mouse model. *Blood*. (2014) 124:3092–100. doi: 10.1182/blood-2014-04-566687
12. Gothe F, Stremenova Spegarova J, Hatton CF, Griffin H, Sargent T, Cowley SA, et al. Aberrant inflammatory responses to type I interferon in STAT2 or IRF9 deficiency. *J Allergy Clin Immunol*. (2022) 150:955–64.e16. doi: 10.1016/j.jaci.2022.01.026
13. Zhang L, Kuca K, You L, Zhao Y, Musilek K, Nepovimova E, et al. Signal transducer and activator of transcription 3 signaling in tumor immune evasion. *Pharmacol Ther*. (2022) 230:107969. doi: 10.1016/j.pharmthera.2021.107969
14. Liao N, Lakyushin A, Lucet IS, Murphy JM, Yao S, Whitlock E, et al. The molecular basis of JAK/STAT inhibition by SOCS1. *Nat Commun*. (2018) 9:1558. doi: 10.1038/s41467-018-04013-1
15. Kanno H, Matsumoto S, Yoshizumi T, Nakahara K, Kubo A, Murata H, et al. Role of SOCS and VHL proteins in neuronal differentiation and development. *Int J Mol Sci*. (2023) 24:3880. doi: 10.3390/ijms24043880
16. Pike KA, Tremblay ML. Protein tyrosine phosphatases: regulators of CD4 T cells in inflammatory bowel disease. *Front Immunol*. (2018) 9:2504. doi: 10.3389/fimmu.2018.02504
17. Gravina AG, Panarese I, Trotta MC, D'Amico M, Pellegrino R, Ferraraccio F, et al. Melanocortin 3,5 receptors immunohistochemical expression in colonic mucosa of inflammatory bowel disease patients: A matter of disease activity. *World J Gastroenterol*. (2024) 30:1132–42. doi: 10.3747/wjg.v30.i9.1132
18. Xu Y, Yan J, Tao Y, Qian X, Zhang C, Yin L, et al. Pituitary hormone α -MSH promotes tumor-induced myelopoiesis and immunosuppression. *Science*. (2022) 377:1085–91. doi: 10.1126/science.abj2674
19. McDonald T, Muhammad F, Peters K, Lee DJ. Combined deficiency of the melanocortin 5 receptor and adenosine 2A receptor unexpectedly provides resistance to autoimmune disease in a CD8(+) T cell-dependent manner. *Front Immunol*. (2021) 12:742154. doi: 10.3389/fimmu.2021.742154
20. Gravina AG, Pellegrino R, Durante T, Palladino G, Imperio G, D'Amico G, et al. The melanocortin system in inflammatory bowel diseases: insights into its mechanisms and therapeutic potentials. *Cells*. (2023) 12:1889. doi: 10.3390/cells12141889
21. Kleiner S, Rüdrieh U, Gehring M, Loser K, Eiz-Vesper B, Noubissi Nzeteu GA, et al. Human basophils release the anti-inflammatory cytokine IL-10 following stimulation with α -melanocyte-stimulating hormone. *J Allergy Clin Immunol*. (2021) 147:1521–3.e3. doi: 10.1016/j.jaci.2020.12.645
22. Ottani A, Giuliani D, Neri L, Calevro A, Canali F, Vandini E, et al. NDP- α -MSH attenuates heart and liver responses to myocardial reperfusion via the vagus nerve and JAK/ERK/STAT signaling. *Eur J Pharmacol*. (2015) 769:22–32. doi: 10.1016/j.ejphar.2015.10.022
23. Ottani A, Neri L, Canali F, Calevro A, Rossi R, Cappelli G, et al. Protective effects of the melanocortin analog NDP- α -MSH in rats undergoing cardiac arrest. *Eur J Pharmacol*. (2014) 745:108–16. doi: 10.1016/j.ejphar.2014.10.022
24. Ottani A, Galantucci M, Ardimento E, Neri L, Canali F, Calevro A, et al. Modulation of the JAK/ERK/STAT signaling in melanocortin-induced inhibition of local and systemic responses to myocardial ischemia/reperfusion. *Pharmacol Res*. (2013) 72:1–8. doi: 10.1016/j.phrs.2013.03.005
25. Buggy JJ. Binding of alpha-melanocyte-stimulating hormone to its G-protein-coupled receptor on B-lymphocytes activates the Jak/STAT pathway. *Biochem J*. (1998) 331:211–6. doi: 10.1042/bj3310211
26. Grievnikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, et al. IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell*. (2009) 15:103–13. doi: 10.1016/j.ccr.2009.01.001
27. Lee H, Herrmann A, Deng JH, Kujawski M, Niu G, Li Z, et al. Persistently activated Stat3 maintains constitutive NF-kappaB activity in tumors. *Cancer Cell*. (2009) 15:283–93. doi: 10.1016/j.ccr.2009.02.015
28. Wang H, Wei W, Zhang JP, Song Z, Li Y, Xiao W, et al. A novel model of alternative NF-kB pathway activation in anaplastic large cell lymphoma. *Leukemia*. (2021) 35:1976–89. doi: 10.1038/s41375-020-01088-y
29. Heinrich PC, Behrmann I, Haan S, Hermanns HM, Müller-Newen G, Schaper F. Principles of interleukin (IL)-6-type cytokine signalling and its regulation. *Biochem J*. (2003) 374:1–20. doi: 10.1042/BJ20030407
30. Larabi A, Barnich N, Nguyen H. New insights into the interplay between autophagy, gut microbiota and inflammatory responses in IBD. *Autophagy*. (2020) 16:38–51. doi: 10.1080/15548627.2019.1635384
31. Liu Z, Hu K, Chen YS, Huang YJ, Hu Q, Zeng W, et al. JAK2/STAT3 inhibition attenuates intestinal ischemia-reperfusion injury via promoting autophagy: *in vitro* and *in vivo* study. *Mol Biol Rep*. (2022) 49:2857–67. doi: 10.1007/s11033-021-07099-x
32. Chang YP, Tsai CC, Huang WC, Wang CY, Chen CL, Lin YS, et al. Autophagy facilitates IFN-gamma-induced Jak2-STAT1 activation and cellular inflammation. *J Biol Chem*. (2010) 285:28715–22. doi: 10.1074/jbc.M110.133355
33. Singh A, Goyal MK, Midha V, Mahajan R, Kaur K, Gupta YK, et al. Tofacitinib in acute severe ulcerative colitis (TACOS): A randomized controlled trial. *Am J Gastroenterol*. (2024) 119:1365–72. doi: 10.14309/ajg.0000000000002635
34. Niehues H, Rösler B, van der Krieken DA, van Vlijmen-Willems I, Rodijk-Olthuis D, Peppelman M, et al. STAT1 gain-of-function compromises skin host defense in the context of IFN- γ signaling. *J Allergy Clin Immunol*. (2019) 143:1626–9.e5. doi: 10.1016/j.jaci.2018.11.033
35. Zhao Y, Luan H, Jiang H, Xu Y, Wu X, Zhang Y, et al. Gegen Qinlian decoction relieved DSS-induced ulcerative colitis in mice by modulating Th17/Treg cell homeostasis via suppressing IL-6/JAK2/STAT3 signaling. *Phytomedicine*. (2021) 84:153519. doi: 10.1016/j.phymed.2021.153519
36. Planell N, Lozano JJ, Mora-Buch R, Masamunt MC, Jimeno M, Ordás I, et al. Transcriptional analysis of the intestinal mucosa of patients with ulcerative colitis in remission reveals lasting epithelial cell alterations. *Gut*. (2013) 62:967–76. doi: 10.1136/gutjnl-2012-303333
37. Can G, Tezel A, Gürkan H, Tozkır H, Ünsal G, Soylu AR, et al. Investigation of IL23R, JAK2, and STAT3 gene polymorphisms and gene-gene interactions in Crohn's disease and ulcerative colitis in a Turkish population. *Turk J Gastroenterol*. (2016) 27:525–36. doi: 10.5152/tjg.2016.16327
38. Zhang Q, Zhao K, Shen Q, Han Y, Gu Y, Li X, et al. Tet2 is required to resolve inflammation by recruiting Hdac2 to specifically repress IL-6. *Nature*. (2015) 525:389–93. doi: 10.1038/nature15252
39. Chaudhuri D, Sasaki K, Karkar A, Sharif S, Lewis K, Mammen MJ, et al. Corticosteroids in COVID-19 and non-COVID-19 ARDS: a systematic review and meta-analysis. *Intensive Care Med*. (2021) 47:521–37. doi: 10.1007/s00134-021-06394-2
40. Mencarelli A, Vacca M, Khameneh HJ, Acerbi E, Tay A, Zolezzi F, et al. Calcineurin B in CD4(+) T cells prevents autoimmune colitis by negatively regulating the JAK/STAT pathway. *Front Immunol*. (2018) 9:261. doi: 10.3389/fimmu.2018.00261
41. Uhlig HH, McKenzie BS, Hue S, Thompson C, Joyce-Shaikh B, Stepankova R, et al. Differential activity of IL-12 and IL-23 in mucosal and systemic innate immune pathology. *Immunity*. (2006) 25:309–18. doi: 10.1016/j.immuni.2006.05.017
42. Souza HS, Tortori CJ, Castelo-Branco MT, Carvalho AT, Margallo VS, Delgado CF, et al. Apoptosis in the intestinal mucosa of patients with inflammatory bowel disease: evidence of altered expression of FasL and perforin cytotoxic pathways. *Int J Colorectal Dis*. (2005) 20:277–86. doi: 10.1007/s00384-004-0639-8
43. Luan XZ, Yuan SX, Chen XJ, Zhou YR, Tang H, Li JQ, et al. ODF3B affects the proliferation and apoptosis of glioma via the JAK/STAT pathway. *Am J Cancer Res*. (2024) 14:1419–32. doi: 10.62347/GHKF1995
44. Li Y, Tian Y, Zhu W, Gong J, Zhang W, Yu C, et al. Triptolide induces suppressor of cytokine signaling-3 expression and promotes lamina propria mononuclear cells apoptosis in Crohn's colitis. *Int Immunopharmacol*. (2013) 16:268–74. doi: 10.1016/j.intimp.2013.04.018
45. Yao D, Zhou Z, Wang P, Zheng L, Huang Y, Duan Y, et al. MiR-125-5p/IL-6R axis regulates macrophage inflammatory response and intestinal epithelial cell apoptosis in ulcerative colitis through JAK1/STAT3 and NF-kB pathway. *Cell Cycle*. (2021) 20:2547–64. doi: 10.1080/15384101.2021.1995128
46. Koukos G, Polytrachou C, Kaplan JL, Morley-Fletcher A, Gras-Miralles B, Kokkotou E, et al. MicroRNA-124 regulates STAT3 expression and is down-regulated in colon tissues of pediatric patients with ulcerative colitis. *Gastroenterology*. (2013) 145:842–52.e2. doi: 10.1053/j.gastro.2013.07.001
47. Luo Y, Yu MH, Yan YR, Zhou Y, Qin SL, Huang YZ, et al. Rab27A promotes cellular apoptosis and ROS production by regulating the miRNA-124-3p/STAT3/RelA signalling pathway in ulcerative colitis. *J Cell Mol Med*. (2020) 24:11330–42. doi: 10.1111/jcmm.15726
48. Rossini V, Zhurina D, Radulovic K, Manta C, Walther P, Riedel CU, et al. CX3CR1⁺ cells facilitate the activation of CD4 T cells in the colonic lamina propria during antigen-driven colitis. *Mucosal Immunol*. (2014) 7:533–48. doi: 10.1038/mi.2013.70

49. Koscsó B, Kurapati S, Rodrigues RR, Nedjic J, Gowda K, Shin C, et al. Gut-resident CX3CR1(hi) macrophages induce tertiary lymphoid structures and IgA response *in situ*. *Sci Immunol*. (2020) 5:eax0062. doi: 10.1126/sciimmunol.aax0062
50. Uo M, Hisamatsu T, Miyoshi J, Kaito D, Yoneno K, Kitazume MT, et al. Mucosal CXCR4+ IgG plasma cells contribute to the pathogenesis of human ulcerative colitis through FcγR-mediated CD14 macrophage activation. *Gut*. (2013) 62:1734–44. doi: 10.1136/gutjnl-2012-303063
51. Na YR, Stakenborg M, Seok SH, Matteoli G. Macrophages in intestinal inflammation and resolution: a potential therapeutic target in IBD. *Nat Rev Gastroenterol Hepatol*. (2019) 16:531–43. doi: 10.1038/s41575-019-0172-4
52. Shaw TN, Houston SA, Wemyss K, Bridgeman HM, Barbera TA, Zangerle-Murray T, et al. Tissue-resident macrophages in the intestine are long lived and defined by Tim-4 and CD4 expression. *J Exp Med*. (2018) 215:1507–18. doi: 10.1084/jem.20180019
53. Sheng J, Ruedl C, Karjalainen K. Most tissue-resident macrophages except microglia are derived from fetal hematopoietic stem cells. *Immunity*. (2015) 43:382–93. doi: 10.1016/j.immuni.2015.07.016
54. Schulz C, Gomez Perdiguerio E, Chorro L, Szabo-Rogers H, Cagnard N, Kierdorf K, et al. A lineage of myeloid cells independent of Myb and hematopoietic stem cells. *Science*. (2012) 336:86–90. doi: 10.1126/science.1219179
55. Hedl M, Proctor DD, Abraham C. JAK2 disease-risk variants are gain of function and JAK signaling threshold determines innate receptor-induced proinflammatory cytokine secretion in macrophages. *J Immunol*. (2016) 197:3695–704. doi: 10.4049/jimmunol.1600845
56. Liu Y, Wang L, Li S, Zhang T, Chen C, Hu J, et al. Mechanical stimulation improves rotator cuff tendon-bone healing via activating IL-4/JAK/STAT signaling pathway mediated macrophage M2 polarization. *J Orthop Translat*. (2022) 37:78–88. doi: 10.1016/j.jot.2022.08.008
57. Wang J, Gao H, Xie Y, Wang P, Li Y, Zhao J, et al. Lycium barbarum polysaccharide alleviates dextran sodium sulfate-induced inflammatory bowel disease by regulating M1/M2 macrophage polarization via the STAT1 and STAT6 pathways. *Front Pharmacol*. (2023) 14:1044576. doi: 10.3389/fphar.2023.1044576
58. Zhang H, Zhao B, Wei H, Zeng H, Sheng D, Zhang Y. Cucurbitacin B controls M2 macrophage polarization to suppresses metastasis via targeting JAK-2/STAT3 signalling pathway in colorectal cancer. *J Ethnopharmacol*. (2022) 287:114915. doi: 10.1016/j.jep.2021.114915
59. Lu Z, Xiong W, Xiao S, Lin Y, Yu K, Yue G, et al. Huanglian Jiedu Decoction ameliorates DSS-induced colitis in mice via the JAK2/STAT3 signalling pathway. *Chin Med*. (2020) 15:45. doi: 10.1186/s13020-020-00327-9
60. Kamada N, Hisamatsu T, Okamoto S, Chinen H, Kobayashi T, Sato T, et al. Unique CD14 intestinal macrophages contribute to the pathogenesis of Crohn disease via IL-23/IFN-γ-axis. *J Clin Invest*. (2008) 118:2269–80. doi: 10.1172/JCI34610
61. Nakanishi Y, Sato T, Takahashi K, Ohteki T. IFN-γ-dependent epigenetic regulation instructs colitogenic monocyte/macrophage lineage differentiation. *vivo. Mucosal Immunol*. (2018) 11:871–80. doi: 10.1038/mi.2017.104
62. Lawrence T, Natoli G. Transcriptional regulation of macrophage polarization: enabling diversity with identity. *Nat Rev Immunol*. (2011) 11:750–61. doi: 10.1038/nri3088
63. Long J, Liu XK, Kang ZP, Wang MX, Zhao HM, Huang JQ, et al. Ginsenoside Rg1 ameliorates experimental colitis by regulating the balance of M1/M2 macrophage polarization and the homeostasis of intestinal flora. *Eur J Pharmacol*. (2022) 917:174742. doi: 10.1016/j.ejphar.2022.174742
64. Zhou X, Li W, Wang S, Zhang P, Wang Q, Xiao J, et al. YAP aggravates inflammatory bowel disease by regulating M1/M2 macrophage polarization and gut microbial homeostasis. *Cell Rep*. (2019) 27:1176–89.e5. doi: 10.1016/j.celrep.2019.03.028
65. Dharmasiri S, Garrido-Martin EM, Harris RJ, Bateman AC, Collins JE, Cummings J, et al. Human intestinal macrophages are involved in the pathology of both ulcerative colitis and crohn disease. *Inflammation Bowel Dis*. (2021) 27:1641–52. doi: 10.1093/ibd/izab029
66. Cheng Y, Li J, Wang L, Wu X, Li Y, Xu M, et al. Eriocalyxin B ameliorated Crohn's disease-like colitis by restricting M1 macrophage polarization through JAK2/STAT1 signalling. *Eur J Pharmacol*. (2023) 954:175876. doi: 10.1016/j.ejphar.2023.175876
67. Ding N, Wang Y, Dou C, Liu F, Guan G, Wei K, et al. Physalin D regulates macrophage M1/M2 polarization via the STAT1/6 pathway. *J Cell Physiol*. (2019) 234:8788–96. doi: 10.1002/jcp.27537
68. Iwata H, Goettsch C, Sharma A, Ricchiuto P, Goh WW, Halu A, et al. PARP9 and PARP14 cross-regulate macrophage activation via STAT1 ADP-ribosylation. *Nat Commun*. (2016) 7:12849. doi: 10.1038/ncomms12849
69. Zhu X, Guo Q, Zou J, Wang B, Zhang Z, Wei R, et al. MiR-19a-3p suppresses M1 macrophage polarization by inhibiting STAT1/IRF1 pathway. *Front Pharmacol*. (2021) 12:614044. doi: 10.3389/fphar.2021.614044
70. Qin H, Yeh WI, De Sarno P, Holdbrooks AT, Liu Y, Muldowney MT, et al. Signal transducer and activator of transcription-3/suppressor of cytokine signaling-3 (STAT3/SOCS3) axis in myeloid cells regulates neuroinflammation. *Proc Natl Acad Sci U.S.A.* (2012) 109:5004–9. doi: 10.1073/pnas.1117218109
71. Zheng Y, Ji S, Li X, Wen L. Qijia rougan formula ameliorates ECM deposition in hepatic fibrosis by regulating the JAK1/STAT6-microRNA-23a feedback loop in macrophage M2 polarization. *BioMed Pharmacother*. (2023) 168:115794. doi: 10.1016/j.biopha.2023.115794
72. Guo J, Tang X, Deng P, Hui H, Chen B, An J, et al. Interleukin-4 from curcumin-activated OECs emerges as a central modulator for increasing M2 polarization of microglia/macrophage in OEC anti-inflammatory activity for functional repair of spinal cord injury. *Cell Commun Signal*. (2024) 22:162. doi: 10.1186/s12964-024-01539-4
73. He L, Du J, Chen Y, Liu C, Zhou M, Adhikari S, et al. Renin-angiotensin system promotes colonic inflammation by inducing T(H)17 activation via JAK2/STAT pathway. *Am J Physiol Gastrointest Liver Physiol*. (2019) 316:G774–774G784. doi: 10.1152/ajpgi.00053.2019
74. Li HB, Tong J, Zhu S, Batista PJ, Duffy EE, Zhao J, et al. m(6)A mRNA methylation controls T cell homeostasis by targeting the IL-7/STAT5/SOCS pathways. *Nature*. (2017) 548:338–42. doi: 10.1038/nature23450
75. Fan Q, Li M, Zhao W, Zhang K, Li M, Li W. Hyper α2,6-sialylation promotes CD4(+) T-cell activation and induces the occurrence of ulcerative colitis. *Adv Sci (Weinh)*. (2023) 10:e2302607. doi: 10.1002/adv.2032607
76. Centifanto-Fitzgerald Y, Caldwell DR, Yates F. Herpes simplex virus: recurrent and nonrecurrent strains. *Proc Soc Exp Biol Med*. (1987) 185:484–92. doi: 10.3181/00379727-185-42574
77. Bsat M, Chapuy L, Rubio M, Wassef R, Richard C, Schwenter F, et al. Differential pathogenic th17 profile in mesenteric lymph nodes of crohn's disease and ulcerative colitis patients. *Front Immunol*. (2019) 10:1177. doi: 10.3389/fimmu.2019.01177
78. Ueno A, Jeffery L, Kobayashi T, Hibi T, Ghosh S, Jijon H. Th17 plasticity and its relevance to inflammatory bowel disease. *J Autoimmun*. (2018) 87:38–49. doi: 10.1016/j.jaut.2017.12.004
79. Hori S, Nomura T, Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. *Science*. (2003) 299:1057–61. doi: 10.1126/science.1079490
80. Ivanov II, Zhou L, Littman DR. Transcriptional regulation of Th17 cell differentiation. *Semin Immunol*. (2007) 19:409–17. doi: 10.1016/j.smim.2007.10.011
81. Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, et al. STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. *J Biol Chem*. (2007) 282:9358–63. doi: 10.1074/jbc.C600321200
82. Hop HT, Huy T, Reyes A, Arayan LT, Vu SH, Min W, et al. Interleukin 6 promotes brucella abortus clearance by controlling bactericidal activity of macrophages and CD8(+) T cell differentiation. *Infect Immun*. (2019) 87:e00431–19. doi: 10.1128/IAI.00431-19
83. Schiavinato J, Haddad R, Saldanha-Araujo F, Baiocchi J, Araujo AG, Santos Scheucher P, et al. TGF-β/αRA-induced Tregs express a selected set of microRNAs involved in the repression of transcripts related to Th17 differentiation. *Sci Rep*. (2017) 7:3627. doi: 10.1038/s41598-017-03456-8
84. Huang X, Lin R, Liu H, Dai M, Guo J, Hui W, et al. Resatorvid (TAK-242) ameliorates ulcerative colitis by modulating macrophage polarization and T helper cell balance via TLR4/JAK2/STAT3 signaling pathway. *Inflammation*. (2024) 47:2108–28. doi: 10.1007/s10753-024-02028-z
85. Xu KX, Xue MG, Li Z, Ye BC, Zhang B. Recent progress on feasible strategies for arbutin production. *Front Bioeng Biotechnol*. (2022) 10:914280. doi: 10.3389/fbioe.2022.914280
86. Wang L, Feng Y, Wang J, Luo T, Wang X, Wu M, et al. Arbutin ameliorates murine colitis by inhibiting JAK2 signaling pathway. *Front Pharmacol*. (2021) 12:683818. doi: 10.3389/fphar.2021.683818
87. Qin D, Liu J, Guo W, Ju T, Fu S, Liu D, et al. Arbutin alleviates intestinal colitis by regulating neutrophil extracellular traps formation and microbiota composition. *Phytomedicine*. (2024) 130:155741. doi: 10.1016/j.phymed.2024.155741
88. Yuan H, Jiang S, Liu Y, Daniyal M, Jian Y, Peng C, et al. The flower head of Chrysanthemum morifolium Ramat. (Juhua): A paradigm of flowers serving as Chinese dietary herbal medicine. *J Ethnopharmacol*. (2020) 261:113043. doi: 10.1016/j.jep.2020.113043
89. Zheng C, Dong Q, Chen H, Cong Q, Ding K. Structural characterization of a polysaccharide from Chrysanthemum morifolium flowers and its antioxidant activity. *Carbohydr Polym*. (2015) 130:113–21. doi: 10.1016/j.carbpol.2015.05.004
90. Hang H, Chen R, Wang C, Sun Y, Du D. A review of the extraction processes and biological characteristics of Chrysanthemum polysaccharides. *Int J Biol Macromol*. (2025) 285:138224. doi: 10.1016/j.ijbiomac.2024.138224
91. Zheng C, Dong Q, Du Z, Wang P, Ding K. Structural elucidation of a polysaccharide from Chrysanthemum morifolium flowers with anti-angiogenic activity. *Int J Biol Macromol*. (2015) 79:674–80. doi: 10.1016/j.ijbiomac.2015.04.026
92. Xiong Q, Luo G, Zheng F, Wu K, Yang H, Chen L, et al. Structural characterization and evaluation the elicitors activity of polysaccharides from Chrysanthemum indicum. *Carbohydr Polym*. (2021) 263:117994. doi: 10.1016/j.carbpol.2021.117994
93. Guo H, Yuan Q, Fu Y, Liu W, Su YH, Liu H, et al. Extraction optimization and effects of extraction methods on the chemical structures and antioxidant activities of polysaccharides from snow chrysanthemum (*Coreopsis tinctoria*). *Polymers (Basel)*. (2019) 11:215. doi: 10.3390/polym11020215

94. Tao JH, Duan JA, Zhang W, Jiang S, Guo JM, Wei DD. Polysaccharides From *Chrysanthemum morifolium* Ramat Ameliorate Colitis Rats via Regulation of the Metabolic Profiling and NF- κ B/TLR4 and IL-6/JAK2/STAT3 Signaling Pathways. *Front Pharmacol.* (2018) 9:746. doi: 10.3389/fphar.2018.00746
95. Feng Z, Ye W, Feng L. Bioactives and metabolites of *Tetragium hemsleyanum* root extract alleviate DSS-induced ulcerative colitis by targeting the SYK protein in the B cell receptor signaling pathway. *J Ethnopharmacol.* (2024) 322:117563. doi: 10.1016/j.jep.2023.117563
96. Fu S, Bao X, Mao Z, Lv Y, Zhu B, Chen Y, et al. Tetragium hemsleyanum polysaccharide ameliorates cytokine storm syndrome via the IFN- γ -JAK2/STAT pathway. *Int J Biol Macromol.* (2024) 275:133427. doi: 10.1016/j.ijbiomac.2024.133427
97. Bao X, Tang Y, Lv Y, Fu S, Yang L, Chen Y, et al. Tetragium hemsleyanum polysaccharide ameliorated ulcerative colitis by remodeling intestinal mucosal barrier function via regulating the SOCS1/JAK2/STAT3 pathway. *Int Immunopharmacol.* (2024) 137:112404. doi: 10.1016/j.intimp.2024.112404
98. Lin Y, Lv Y, Mao Z, Chen X, Chen Y, Zhu B, et al. Polysaccharides from *Tetragium Hemsleyanum* Diels et Gilg ameliorated inflammatory bowel disease by rebuilding the intestinal mucosal barrier and inhibiting inflammation through the SCFA-GPR41/43 signaling pathway. *Int J Biol Macromol.* (2023) 250:126167. doi: 10.1016/j.ijbiomac.2023.126167
99. Korkina L, Suprun M, Petrova A, Mikhal'chik E, Luci A, De Luca C. The protective and healing effects of a natural antioxidant formulation based on ubiquinol and Aloe vera against dextran sulfate-induced ulcerative colitis in rats. *Biofactors.* (2003) 18:255–64. doi: 10.1002/biof.5520180228
100. Xu L, Zheng X, Wang Y, Fan Q, Zhang M, Li R, et al. Berberine protects acute liver failure in mice through inhibiting inflammation and mitochondria-dependent apoptosis. *Eur J Pharmacol.* (2018) 819:161–8. doi: 10.1016/j.ejphar.2017.11.013
101. Jia X, Shao W, Tian S. Berberine alleviates myocardial ischemia-reperfusion injury by inhibiting inflammatory response and oxidative stress: the key function of miR-26b-5p-mediated PTGS2/MAPK signal transduction. *Pharm Biol.* (2022) 60:652–63. doi: 10.1080/13880209.2022.2048029
102. Sun Q, Yang H, Liu M, Ren S, Zhao H, Ming T, et al. Berberine suppresses colorectal cancer by regulation of Hedgehog signaling pathway activity and gut microbiota. *Phytomedicine.* (2022) 103:154227. doi: 10.1016/j.phymed.2022.154227
103. Li H, Fan C, Lu H, Feng C, He P, Yang X, et al. Protective role of berberine on ulcerative colitis through modulating enteric glial cells-intestinal epithelial cells-immune cells interactions. *Acta Pharm Sin B.* (2020) 10:447–61. doi: 10.1016/j.apsb.2019.08.006
104. Xu L, Zhang Y, Xue X, Liu J, Li ZS, Yang GY, et al. A phase I trial of berberine in chinese with ulcerative colitis. *Cancer Prev Res (Phila).* (2020) 13:117–26. doi: 10.1158/1940-6207.CAPR-19-0258
105. Gravina AG, Pellegrino R, Palladino G, Coppola A, Brandimarte G, Tuccillo C, et al. *Hericum erinaceus*, in combination with natural flavonoid/alkaloid and B(3)/B(8) vitamins, can improve inflammatory burden in inflammatory bowel diseases tissue: an ex vivo study. *Front Immunol.* (2023) 14:1215329. doi: 10.3389/fimmu.2023.1215329
106. Dong Y, Fan H, Zhang Z, Jiang F, Li M, Zhou H, et al. Berberine ameliorates DSS-induced intestinal mucosal barrier dysfunction through microbiota-dependence and Wnt/ β -catenin pathway. *Int J Biol Sci.* (2022) 18:1381–97. doi: 10.7150/ijbs.65476
107. Jing W, Dong S, Luo X, Liu J, Wei B, Du W, et al. Berberine improves colitis by triggering AhR activation by microbial tryptophan catabolites. *Pharmacol Res.* (2021) 164:105358. doi: 10.1016/j.phrs.2020.105358
108. Sun X, Zhang Y, Cheng G, Zhu T, Zhang Z, Xiong L, et al. Berberine improves DSS-induced colitis in mice by modulating the fecal-bacteria-related bile acid metabolism. *BioMed Pharmacother.* (2023) 167:115430. doi: 10.1016/j.bioph.2023.115430
109. Zhu L, Gu P, Shen H. Protective effects of berberine hydrochloride on DSS-induced ulcerative colitis in rats. *Int Immunopharmacol.* (2019) 68:242–51. doi: 10.1016/j.intimp.2018.12.036
110. Hermanns HM. Oncostatin M. and interleukin-31: Cytokines, receptors, signal transduction and physiology. *Cytokine Growth Factor Rev.* (2015) 26:545–58. doi: 10.1016/j.cytogfr.2015.07.006
111. Ye Y, Fu Y, Lin C, Shen Y, Yu Q, Yao X, et al. Oncostatin M induces IFITM1 expression to inhibit hepatitis B virus replication via JAK-STAT signaling. *Cell Mol Gastroenterol Hepatol.* (2024) 17:219–35. doi: 10.1016/j.jcmgh.2023.10.003
112. Li H, Feng C, Fan C, Yang Y, Yang X, Lu H, et al. Intervention of oncostatin M-driven mucosal inflammation by berberine exerts therapeutic property in chronic ulcerative colitis. *Cell Death Dis.* (2020) 11:271. doi: 10.1038/s41419-020-2470-8
113. Xiong K, Deng J, Yue T, Hu W, Zeng X, Yang T, et al. Berberine promotes M2 macrophage polarisation through the IL-4-STAT6 signalling pathway in ulcerative colitis treatment. *Heliyon.* (2023) 9:e14176. doi: 10.1016/j.heliyon.2023.e14176
114. Lin Y, He F, Wu L, Xu Y, Du Q. Matrine exerts pharmacological effects through multiple signaling pathways: A comprehensive review. *Drug Des Devel Ther.* (2022) 16:533–69. doi: 10.2147/DDDT.S349678
115. Mao N, Yu Y, He J, Yang Y, Liu Z, Lu Y, et al. Matrine ameliorates DSS-induced colitis by suppressing inflammation, modulating oxidative stress and remodeling the gut microbiota. *Int J Mol Sci.* (2024) 25:6613. doi: 10.3390/ijms25126613
116. Chen A, Fang D, Ren Y, Wang Z. Matrine protects colon mucosal epithelial cells against inflammation and apoptosis via the Janus kinase 2/signal transducer and activator of transcription 3 pathway. *Bioengineered.* (2022) 13:6490–9. doi: 10.1080/21655979.2022.2031676
117. Elhennawy MG, Abdelaleem EA, Zaki AA, Mohamed WR. Cinnamaldehyde and hesperetin attenuate TNBS-induced ulcerative colitis in rats through modulation of the JAK2/STAT3/SOCS3 pathway. *J Biochem Mol Toxicol.* (2021) 35:e22730. doi: 10.1002/jbt.22730
118. Sadeghi N, Mansoori A, Shayesteh A, Hashemi SJ. The effect of curcumin supplementation on clinical outcomes and inflammatory markers in patients with ulcerative colitis. *Phytother Res.* (2020) 34:1123–33. doi: 10.1002/ptr.6581
119. Zhao HM, Xu R, Huang XY, Cheng SM, Huang MF, Yue HY, et al. Curcumin suppressed activation of dendritic cells via JAK/STAT/SOCS signal in mice with experimental colitis. *Front Pharmacol.* (2016) 7:455. doi: 10.3389/fphar.2016.00455
120. Wei C, Wang JY, Xiong F, Wu BH, Luo MH, Yu ZC, et al. Curcumin ameliorates DSS-induced colitis in mice by regulating the Treg/Th17 signaling pathway. *Mol Med Rep.* (2021) 23:34. doi: 10.3892/mmr.2020.11672
121. Yao H, Wang F, Chong H, Wang J, Bai Y, Du M, et al. A curcumin-modified coordination polymers with ROS scavenging and macrophage phenotype regulating properties for efficient ulcerative colitis treatment. *Adv Sci (Weinh).* (2023) 10:e2300601. doi: 10.1002/advs.202300601
122. Zhong YB, Kang ZP, Zhou BG, Wang HY, Long J, Zhou W, et al. Curcumin regulated the homeostasis of memory T cell and ameliorated dextran sulfate sodium-induced experimental colitis. *Front Pharmacol.* (2020) 11:630244. doi: 10.3389/fphar.2020.630244
123. Liu L, Liu YL, Liu GX, Chen X, Yang K, Yang YX, et al. Curcumin ameliorates dextran sulfate sodium-induced experimental colitis by blocking STAT3 signaling pathway. *Int Immunopharmacol.* (2013) 17:314–20. doi: 10.1016/j.intimp.2013.06.020
124. Yao J, Wang JY, Liu L, Li YX, Xun AY, Zeng WS, et al. Anti-oxidant effects of resveratrol on mice with DSS-induced ulcerative colitis. *Arch Med Res.* (2010) 41:288–94. doi: 10.1016/j.arcmed.2010.05.002
125. Britton RG, Kovoor C, Brown K. Direct molecular targets of resveratrol: identifying key interactions to unlock complex mechanisms. *Ann N Y Acad Sci.* (2015) 1348:124–33. doi: 10.1111/nyas.12796
126. Xu J, Liu D, Niu H, Zhu G, Xu Y, Ye D, et al. Resveratrol reverses Doxorubicin resistance by inhibiting epithelial-mesenchymal transition (EMT) through modulating PTEN/Akt signaling pathway in gastric cancer. *J Exp Clin Cancer Res.* (2017) 36:19. doi: 10.1186/s13046-016-0487-8
127. Jing Y, Hu T, Yuan J, Liu Z, Tao M, Ou M, et al. Resveratrol protects against postmenopausal atherosclerosis progression through reducing PCSK9 expression via the regulation of the ER α -mediated signaling pathway. *Biochem Pharmacol.* (2023) 211:115541. doi: 10.1016/j.bcp.2023.115541
128. Mahjabeen W, Khan DA, Mirza SA. Role of resveratrol supplementation in regulation of glucose homeostasis, inflammation and oxidative stress in patients with diabetes mellitus type 2: A randomized, placebo-controlled trial. *Complement Ther Med.* (2022) 66:102819. doi: 10.1016/j.ctim.2022.102819
129. Chen JY, Zhu Q, Zhang S, OuYang D, Lu JH. Resveratrol in experimental Alzheimer's disease models: A systematic review of preclinical studies. *Pharmacol Res.* (2019) 150:104476. doi: 10.1016/j.phrs.2019.104476
130. Yu B, Wang Y, Tan Z, Hong Z, Yao L, Huang S, et al. Resveratrol ameliorates DSS-induced ulcerative colitis by acting on mouse gut microbiota. *Inflammopharmacology.* (2024) 32:2023–33. doi: 10.1007/s10787-024-01456-5
131. Li F, Han Y, Cai X, Gu M, Sun J, Qi C, et al. Dietary resveratrol attenuated colitis and modulated gut microbiota in dextran sulfate sodium-treated mice. *Food Funct.* (2020) 11:1063–73. doi: 10.1039/c9fo01519a
132. Serra D, Rufino AT, Mendes AF, Almeida LM, Dinis TC. Resveratrol modulates cytokine-induced Jak/STAT activation more efficiently than 5-aminosalicylic acid: an in vitro approach. *PLoS One.* (2014) 9:e109048. doi: 10.1371/journal.pone.0109048
133. Yaqin Z, Kehan W, Yi Z, Najian W, Wei Q, Fei M. Resveratrol alleviates inflammatory bowel disease by inhibiting JAK2/STAT3 pathway activity via the reduction of O-GlcNAcylation of STAT3 in intestinal epithelial cells. *Toxicol Appl Pharmacol.* (2024) 484:116882. doi: 10.1016/j.taap.2024.116882
134. Alrafas HR, Busbee PB, Nagarkatti M, Nagarkatti PS. Resveratrol Downregulates miR-31 to Promote T Regulatory Cells during Prevention of TNBS-Induced Colitis. *Mol Nutr Food Res.* (2020) 64:e1900633. doi: 10.1002/mnfr.201900633
135. Samsami-Kor M, Daryani NE, Asl PR, Hekmatdoost A. Anti-inflammatory effects of resveratrol in patients with ulcerative colitis: A randomized, double-blind, placebo-controlled pilot study. *Arch Med Res.* (2015) 46:280–5. doi: 10.1016/j.arcmed.2015.05.005
136. Pujara N, Wong KY, Qu Z, Wang R, Moniruzzaman M, Rewatkar P, et al. Oral delivery of β -lactoglobulin-nanosphere-encapsulated resveratrol alleviates inflammation in winnie mice with spontaneous ulcerative colitis. *Mol Pharm.* (2021) 18:627–40. doi: 10.1021/acs.molpharmaceut.0c00048
137. Ye H, Wu Q, Zhu Y, Guo C, Zheng X. Ginsenoside Rh2 alleviates dextran sulfate sodium-induced colitis via augmenting TGF β signaling. *Mol Biol Rep.* (2014) 41:5485–90. doi: 10.1007/s11033-014-3422-0
138. Chen X, Xu T, Lv X, Zhang J, Liu S. Ginsenoside Rh2 alleviates ulcerative colitis by regulating the STAT3/miR-214 signaling pathway. *J Ethnopharmacol.* (2021) 274:113997. doi: 10.1016/j.jep.2021.113997

139. Song C, Chen J, Li X, Yang R, Cao X, Zhou L, et al. Limonin ameliorates dextran sulfate sodium-induced chronic colitis in mice by inhibiting PERK-ATF4-CHOP pathway of ER stress and NF- κ B signaling. *Int Immunopharmacol.* (2021) 90:107161. doi: 10.1016/j.intimp.2020.107161
140. Liu S, Zhang S, Lv X, Lu J, Ren C, Zeng Z, et al. Limonin ameliorates ulcerative colitis by regulating STAT3/miR-214 signaling pathway. *Int Immunopharmacol.* (2019) 75:105768. doi: 10.1016/j.intimp.2019.105768
141. Hao L, Mao Y, Park J, Kwon BM, Bae EJ, Park BH. 2'-Hydroxycinnamaldehyde ameliorates imiquimod-induced psoriasiform inflammation by targeting PKM2-STAT3 signaling in mice. *Exp Mol Med.* (2021) 53:875–84. doi: 10.1038/s12276-021-00620-z
142. Yoon YJ, Kim YH, Jin Y, Chi SW, Moon JH, Han DC, et al. 2'-hydroxycinnamaldehyde inhibits cancer cell proliferation and tumor growth by targeting the pyruvate kinase M2. *Cancer Lett.* (2018) 434:42–55. doi: 10.1016/j.canlet.2018.07.015
143. Cheng YH, Chiang CY, Wu CH, Chien CT. 2'-hydroxycinnamaldehyde, a natural product from cinnamon, alleviates ischemia/reperfusion-induced microvascular dysfunction and oxidative damage in rats by upregulating cytosolic BAG3 and nrf2/HO-1. *Int J Mol Sci.* (2024) 25:12962. doi: 10.3390/ijms252312962
144. Yoon YJ, Kim YH, Lee YJ, Choi J, Kim CH, Han DC, et al. 2'-Hydroxycinnamaldehyde inhibits proliferation and induces apoptosis via signal transducer and activator of transcription 3 inactivation and reactive oxygen species generation. *Cancer Sci.* (2019) 110:366–78. doi: 10.1111/cas.13852
145. Chen M, Wei S, Wu X, Xiang Z, Li X, He H, et al. 2'-hydroxycinnamaldehyde alleviates intestinal inflammation by attenuating intestinal mucosal barrier damage via directly inhibiting STAT3. *Inflammation Bowel Dis.* (2024) 30:992–1008. doi: 10.1093/ibd/izad283
146. Fan X, Gao Y, Hua C, Peng L, Ci X. Daphnetin ameliorates PM2.5-induced airway inflammation by inhibiting NLRP3 inflammasome-mediated pyroptosis in CS-exposed mice. *BioMed Pharmacother.* (2023) 165:115047. doi: 10.1016/j.biopha.2023.115047
147. Wang G, Pang J, Hu X, Nie T, Lu X, Li X, et al. Daphnetin: A novel anti-helicobacter pylori agent. *Int J Mol Sci.* (2019) 20:850. doi: 10.3390/ijms20040850
148. Zhang Y, Qu L, Sun Y, Lin Y, Zeng J, He L, et al. Daphnetin contributes to allergen-induced Th2 cytokine expression and type 2-immune responses in atopic dermatitis and asthma. *Food Funct.* (2022) 13:12383–99. doi: 10.1039/d2fo02518c
149. Xia Y, Chen C, Liu Y, Ge G, Dou T, Wang P. Synthesis and structure-activity relationship of daphnetin derivatives as potent antioxidant agents. *Molecules.* (2018) 23:2476. doi: 10.3390/molecules23102476
150. Javed M, Saleem A, Xaveria A, Akhtar MF. Daphnetin: A bioactive natural coumarin with diverse therapeutic potentials. *Front Pharmacol.* (2022) 13:993562. doi: 10.3389/fphar.2022.993562
151. Ji J, Ge X, Chen Y, Zhu B, Wu Q, Zhang J, et al. Daphnetin ameliorates experimental colitis by modulating microbiota composition and T(reg)/T(h)17 balance. *FASEB J.* (2019) 33:9308–22. doi: 10.1096/fj.201802659RR
152. He Z, Liu J, Liu Y. Daphnetin attenuates intestinal inflammation, oxidative stress, and apoptosis in ulcerative colitis via inhibiting REG3A-dependent JAK2/STAT3 signaling pathway. *Environ Toxicol.* (2023) 38:2132–42. doi: 10.1002/tox.23837
153. Fu J, Zang Y, Zhou Y, Chen C, Shao S, Hu M, et al. A novel triptolide derivative ZT01 exerts anti-inflammatory effects by targeting TAK1 to prevent macrophage polarization into pro-inflammatory phenotype. *BioMed Pharmacother.* (2020) 126:110084. doi: 10.1016/j.biopha.2020.110084
154. Fu J, Zang Y, Zhou Y, Chen C, Shao S, Shi G, et al. Exploring a novel triptolide derivative possess anti-colitis effect via regulating T cell differentiation. *Int Immunopharmacol.* (2021) 94:107472. doi: 10.1016/j.intimp.2021.107472